

**EMORY**  
UNIVERSITY**Emory Integrated  
Genomics Core**  
Emory Integrated Core Facilities**EIGC.002\_Appendix M\_QIAGEN QIAamp DNA Mini for Cell  
Culture****Standard Operating Procedure Staff Review Page:**

I have read and understand the procedure listed above.

| Employee name | Date SOP review complete |
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## Standard Operating Procedure Approval Page:

Date Implemented: 23 January 2008

Updated: Ashima Amin  
**Name**

15 June 2020  
**Date**

Supersedes: Ashima Amin  
**Name**

11 April 2016  
**Date**

Annual Review and Approval

Michael Zwick, PhD  
Laboratory Director

**Changes Made:**

061520: Review, Protocols split out into appendixes. Renumbered SOPs. Previously EIGC.003.

***Changes to previous procedures:***

041116: Review, LIMS steps updated

## Introduction

This protocol is for purification of total (genomic, mitochondrial, and viral) DNA from cell culture using a microcentrifuge.

QIAamp DNA Mini Kit provides fast and easy methods for purification of total DNA for reliable PCR and Southern blotting. Total DNA (e.g., genomic, viral, mitochondrial) can be purified from a variety of specimen types, including cell culture.

DNA is purified using a QIAamp spin column, yielding DNA ready for direct amplification or downstream application. The procedure requires no phenol/chloroform extraction or alcohol precipitation and involves very little handling. DNA is eluted in Buffer AE or water. DNA purified using QIAamp Kits is up to 50 kb in size, with fragments of approximately 20–30 kb predominating. DNA of this length denatures completely during thermal cycling and can be amplified with high efficiency. Purified DNA can be stored at 4°C for immediate use or can be safely stored at –20 °C for later use. The purified DNA is free of protein, nucleases, and other contaminants or inhibitors.

## Kit Contents

| QIAamp DNA Kits                             | Blood Mini (50)     | Blood Mini (250)    | Mini (50) | Mini (250) |
|---------------------------------------------|---------------------|---------------------|-----------|------------|
| <b>Catalog no.</b>                          | 51104               | 51106               | 51304     | 51306      |
| <b>Number of preps</b>                      | 50                  | 250                 | 50        | 250        |
| <b>QIAamp Mini Spin Columns</b>             | 50                  | 250                 | 50        | 250        |
| <b>Collection Tubes (2 ml)</b>              | 150                 | 750                 | 150       | 750        |
| <b>Buffer AL<sup>a</sup></b>                | 12 ml               | 2 x 33 ml           | 12 ml     | 2 x 33 ml  |
| <b>Buffer ATL</b>                           | –                   | –                   | 14 ml     | 50 ml      |
| <b>Buffer AW1<sup>a</sup> (concentrate)</b> | 19 ml               | 98 ml               | 19 ml     | 98 ml      |
| <b>Buffer AW2<sup>b</sup> (concentrate)</b> | 13 ml               | 66 ml               | 13 ml     | 66 ml      |
| <b>Buffer AE</b>                            | 15 ml               | 60 ml               | 2 x 15 ml | 128 ml     |
| <b>QIAGEN® Protease<sup>e</sup></b>         | 1 vial <sup>c</sup> | 1 vial <sup>d</sup> | –         | –          |
| <b>Protease Solvent<sup>b</sup></b>         | 1.2 ml              | 5.5 ml              | –         | –          |
| <b>Proteinase K<sup>e</sup></b>             | –                   | –                   | 1.25 ml   | 6 ml       |

<sup>a</sup> Contains chaotropic salt. Not compatible with disinfecting agents containing bleach; see handbook for safety information.

<sup>b</sup> Contains sodium azide as a preservative.

<sup>c</sup> Resuspension volume 1.2 ml.

<sup>d</sup> Resuspension volume 5.5 ml.

<sup>e</sup> QIAGEN® Protease and Proteinase K are stored at 4°C.

## Procedure

### Notes before starting

- No more than  $5 \times 10^6$  cells (diploid) should be used for this protocol.
- Use carrier DNA if the sample contains <10,000 genome equivalents. See handbook for details.
- Heat a water bath or heating block to 56°C.
- All centrifugation steps are carried out at room temperature (15–25°C).
- If a precipitate has formed in Buffer ATL or Buffer AL, dissolve by incubating at 56°C.
- Ensure that Buffer AW1 and Buffer AW2 have been prepared according to the instructions as indicated on the bottle or the table above.
- You will need the following additional equipment and reagents:
  - Phosphate buffered saline (PBS)
  - Microcentrifuge set to room temperature

1. **Determine the number of cells in each sample. Centrifuge up to  $5 \times 10^6$  cells (diploid) at 300 x g for 5 min. Remove and discard the supernatant.**
2. **Briefly vortex the sample to resuspend the cell pellet in the residual media. This step will help with the next suspension step.**
3. **Resuspend the cells in 200 µl PBS.**

$5 \times 10^6$  cells will yield approximately 15–20 µg of DNA.

4. **Add 20 µl QIAGEN Protease (or proteinase K).**
5. **Add 4 µl of an RNase A stock solution (100 mg/ml).**
6. **Add 200 µl Buffer AL to the sample. Mix by pulse-vortexing for 15 s.**

**Note:** Do not add QIAGEN Protease, proteinase K, or RNase A directly to Buffer AL.

7. **Incubate at 56°C for 10 min.**
8. **Briefly centrifuge the 1.5 ml microcentrifuge tube to remove drops from the inside of the lid.**
9. **Add 200 µl ethanol (96–100%) to the sample, and mix again by pulse-vortexing for 15 s. After mixing, briefly centrifuge the 1.5 ml microcentrifuge tube to remove drops from the inside of the lid.**
10. **Carefully apply the mixture from step 6 to the QIAamp Mini spin column (in a 2 ml collection tube) without wetting the rim. Close the cap, and centrifuge at 6000 x g for 1 min. Place the QIAamp Mini spin column in a clean 2 ml collection tube (provided), and discard the tube containing the filtrate.\***

\*Flow-through contains Buffer AL or Buffer AW1 and is therefore not compatible with bleach.

When preparing DNA from buffy coat or lymphocytes, centrifugation at full speed is recommended to avoid clogging.

11. **Carefully open the QIAamp Mini spin column and add 500 µl Buffer AW1 without wetting the rim. Close the cap and centrifuge at 6000 x g for 1 min. Place the QIAamp Mini spin column in a clean 2 ml collection tube (provided), and discard the collection tube containing the filtrate.\***
12. **Carefully open the QIAamp Mini spin column and add 500 µl Buffer AW2 without wetting the rim. Close the cap and centrifuge at full speed (20,000 x g) for 3 min.**
13. **Place the QIAamp Mini spin column in a new 2 ml collection tube (not provided) and discard the old collection tube with the filtrate. Centrifuge at full speed for 1 min.**
14. **Place the QIAamp Mini spin column in a clean 1.5 ml microcentrifuge tube (not provided) and discard the collection tube containing the filtrate. Carefully open the QIAamp Mini spin column and add 200 µl distilled water. Incubate at room temperature (15–25°C) for 5 min and then centrifuge at 6000 x g for 1 min.**
15. **Elute a second time using 100 µl of distilled water, for a total of 300 µl.**

Note: if your sample input is low, you may elute in lower volumes.